Apple *CALCINEURIN B-LIKE PROTEIN10* genes have evolved to be novel targets of miR167s through sequence variation

Ashutosh Kumar and Ananda K. Sarkar*

National Institute of Plant Genome Research, Aruna Asaf Ali Marg, New Delhi 110 067, India

The miR167s and its target *ARF6/8* are relatively conserved among diverse plant species and have been implicated in reproductive and root development in *Arabidopsis*. Here we show that some of the *CNBL* family members have evolved to be targets of miR167s in apple. Despite strong conservation between apple and *Arabidopsis* CNBLs, *AtCNBLs* are not miR167 targets. The sequence variation in apple-miR167a and *MdCNBLs* has created target sites for apple-miR167a in *MdCNBL10s*. Therefore, we suggest that during the course of evolution, natural selection through sequence variation played a crucial role by choosing different targets among plant species for the same miRNA.

Keywords: Arabidopsis thaliana, CNBL10, miR167, Malus domestica (apple), miRNA evolution.

THE diversity among organisms having the same genetic material arises due to the variation in functionality of their molecular mechanism. The molecular mechanism of biological processes involves dynamic network of genes responsible for multiple functions. Besides protein coding genes, small non-protein coding RNAs have recently been shown to be crucial components of gene regulatory network. These small RNAs (sRNAs, 19-24 bp long) regulate diverse developmental processes, including root and shoot growth¹, by transcriptional or posttranscriptional gene silencing mechanisms². The two sRNA classes, microRNAs (miRNAs) and small interfering RNAs (siRNAs) majorly comprise sRNA population in plants³. The miRNAs act either by cleaving target mRNA(s) by pairing with it at complementary sequence to direct post-transcriptional gene silencing (PTGS) or by inhibiting translation of mRNAs^{3,4}. It is generally considered that miRNAs and their target(s) with perfect or nearperfect complementary sites are conserved among various plant species⁵. Similarly, we have recently shown that miR167s, which are required for flower development through negative regulation of AUXIN RESPONSE FACTOR (ARF6/8) transcript, have undergone thorough co-evolution with their target ARF6/8s among diverse plant species^{6,7}. We found a novel non-ARF target of mdm-miR167, CALCINEURIN B-LIKE10 (CNBL10) in apple (Malus domestica), which was experimentally validated through 5' RLM-RACE PCR⁶. Moreover, our study has identified and predicted non-ARF6/8 target in crops such as Oryza sativa, Glycine max, etc⁶. This suggests that sequence variation in both miRNAs and complementary sequence of targets occasionally leads to functional diversification of miRNAs, which is not well addressed. Conventionally, it is assumed that like miRNAs, targets are also conserved among diverse plants species because functionality of miRNAs is depicted through their targets. Changes that occur during the course of evolution in both the mature miRNA sequence and their complementary regions of target genes may lead to alteration in that miRNA-mediated gene regulation and functional diversification. Therefore, for better understanding the regulatory role of a biologically important miRNA, it is prerequisite to understand whether their functional diversification happened through sequence variation in miRNA and/or target sites. In this work, we extended our recent study where we speculated that miRNAs and their targets co-evolved, which played an important role for their functional divergence, and hypothesized the possible cause of the functional divergence of mdm-miR167 in Malus domestica.

The miR167 sequences of Arabidopsis and apple were retrieved from miRNA registry database (miRBase version 21, http://microrna.sanger.ac.uk/). It was found from our recent study, while performing multiple sequence alignment, that the ath-miR167a, b and mdm-miR167b-g have identical sequences, which therefore clustered and were named together as unique miR167-1 (UmiR167-1), whereas ath-miR167c and d have difference in their sequences. Similarly, mdm-miR167a has difference in its sequence and did not cluster with other mdm-miR167s and ath-miR167s, therefore was named as unique miR167-2 (UmiR167-2)⁶. The target for the UmiR167-1 was predicted with the help of psRNATarget tool⁸, as ARF6, which was already reported⁷. The *psRNATarget* tool was used to identify or predict a novel target gene for UmiR167-2, which has been validated through 5' RLM-RACE PCR⁶. The nucleotide and protein sequences of CNBL family members of Arabidopsis and apple were retrieved from TAIR (version 10) and NCBI respectively (Table 1). The nomenclature for the species specific CNBL genes was done by putting the prefix 'At' for Arabidopsis and 'Md' for apple. The gene names CNBL and CBL are synonymous; the entries of protein sequences in UniProt database (http://www.uniprot.org/ uniprot) are indicated as 'CNBL', whereas TAIR indicated these genes as 'CBL'. These sequences were used to check if they were possible target of UmiR167-1 or UmiR167-2 by using psRNATarget tool. Homology between AtCNBLs and MdCNBLs was identified using phylogenetic analysis tool MEGA6 keeping default parameter settings⁹. Further, to reveal critical sequence

^{*}For correspondence. (e-mail: aksarkar@nipgr.ac.in)

CURRENT SCIENCE, VOL. 112, NO. 1, 10 JANUARY 2017

Gene*	Notation*	Protein accession number	Nucleotide accession number	TAIR accession number
AtCBL1	AtCNBL1	AAC26008	AF076251	At4g17615
AtCBL2	AtCNBL2	AAC26009	AF076252	At5g55990
AtCBL3	AtCNBL3	AAC26010	AF076253	At4g26570
AtCBL4	AtCNBL4	AAG28402	AF192886	At5g24270
AtCBL5	AtCNBL5	AAG28401	AF192885	At4g01420
AtCBL6	AtCNBL6	AAG28400	AF192884	At4g16350
AtCBL7	AtCNBL7	AAG10059	AF290434	At4g26560
AtCBL8	AtCNBL8	AAL10300	AF411957	At1g64480
AtCBL9	AtCNBL9	AAL10301	AF411958	At5g47100
AtCBL10	AtCNBL10	AAO72364	AF490607	At4g33000

 Table 1. a, Nucleotide and protein sequences of CNBL family members in Arabidopsis

b, Nucleotide and protein sequences of CNBL family members in apple

Gene*	Notation*	Protein accession number	Gene accession number
MdCBL1	MdCNBL1.1	XP_008373643	LOC103436960
	MdCNBL1.2	XP_008363760	LOC103427483
	MdCNBL1.3	XP_008356465	LOC103420179
MdCBL2	MdCNBL2	XP_008358898	LOC103422620
MdCBL3	MdCNBL3.1	XP_008384609	LOC103447204
	MdCNBL3.2	XP_008355026	LOC103418695
	MdCNBL3.3	XP_008344727	LOC103407603
MdCBL4	MdCNBL4.1	XP_008382769	LOC103445526
	MdCNBL4.2	XP_008376721	LOC103439871
	MdCNBL4.3	XP_008354255	LOC103417881
MdCBL5	N/A	_	_
MdCBL6	N/A	-	-
MdCBL7	MdCNBL7.1	XP_008343454	LOC103406228
	MdCNBL7.2	XP_008341705	LOC103404551
	MdCNBL7.3	XP_008337551	LOC103400660
MdCBL8	N/A		_
MdCBL9	N/A	-	_
MdCBL10	MdCNBL10.1	XP_008391317	LOC103453555
	MdCNBL10.2	XP_008377510	LOC103440590
	MdCNBL10.3	XP_008365025	LOC103428675
	MdCNBL10.4	XP_008357234	LOC103420977

*CNBL and CBL are synonymous; the entries of protein sequences in UniProt database are 'CNBL', whereas genes indicated as 'CBL' in TAIR.

variation between the complementary UmiR167 sequences and their target genes was analysed through $\text{ClustalX}(2.1)^{10}$.

The novelty in the function of plant miRNAs crucially depends on how they evolved, which includes either changes in their functional mature sequences, processing and expression pattern. In apple, the *MdCNBL10* was shown to be the novel target of mdm-miR167a, which is exceptionally processed differently (from 3' end of the precursors), and had undergone sequence diversification⁶. Therefore, it was important to find out the possible diversification in the functionality of these miR167s and *CNBL* genes. To study whether other members of *CNBL* gene family were also targeted by the UmiR167-2, we retrieved all the sequences from *Arabidopsis* and apple. In

148

Arabidopsis, the AtCNBL gene family consisted of 10 members (AtCNBL1-10), whereas in apple, 6 members were found (except MdCNBL5, 6, 8 and 9, which were missing), while some of them had multiple copies (Table 1). Our study identified the homology between AtCNBL and MdCNBL, using the MEGA6 tool (Figure 1 a). We observed that the available members of MdCNBL were orthologous to the same members of AtCNBL, except MdCNBL7.1, 7.2 and 7.3. MdCNBL7.1 and 7.3 clustered with AtCNBL8, whereas MdCNBL7.2 clustered with AtCNBL5 (Figure 1 a). All these members of CNBL gene family were used for checking complementarity with UmiR167-2, using psRNATarget tool, by keeping default parameter settings. Interestingly, the MdCNBL10.2,





Figure 1. *a*, Homology identification between AtCNBL and MdCNBL; *b*, Pairwise alignment between UmiR167-1 and UmiR167-2; *c*, MSA between the targets *MdCNBL10.2*, *10.3* and *10.4* and complementary sequence of UmiR167-2; *d*, MSA among *AtCNBL10.*, *MdCNBL10.2*, *10.3*, *10.4* and complementary sequence of UmiR167-2; *e*, MSA among MdCNBL10.2, MdCNBL10.3, MdCNBL10.4 and AtCNBL10; red-coloured rectangular box represents amino acids encoded by complementary sequence of UmiR167-2.

MdCNBL10.3 and MdCNBL10.4, of which MdCNBL10.1 was closest to AtCNBL10.1; the remaining three members clustered together (Figure 1 a; Table 1). We observed that only MdCNBL10.2, MdCNBL10.3 and MdCNBL10.4 mRNAs were having binding site for UmiR167-2, indicating them as targets of mdm-miR167a. We have recently proved though 5' RLM-RACE PCR that *MdCNBL10.3* mRNA was cleaved by mdm-miR167a (ref. 6). Our detailed sequence analysis and phylogenetic study of MdCNBL10.2, MdCNBL10.3 and MdCNBL10.4 shows perfect sequence identity including miR167 binding sites. Therefore, this suggests that our previously validated target MdCNBL10 included all these three targets. Though AtCNBL10 was orthologue of MdCNBL10, it did not show complementarity with UmiR167-2. Belonging to the same class of miRNA, UmiR167-1 and UmiR167-2 were having different range of targets. The UmiR167-1 was showing complementarity only with AtARF6 and AtARF8, while UmiR167-2 was only complementary to MdCNBL10.2, MdCNBL10.3 and MdCNBL10.4. The sequence alignment of both UmiRNAs with the help of ClustalX(2.1) showed a colossal difference between the

sequences, only showing $\sim 40\%$ identity (Figure 1 b). This suggests that the ability of UmiR167-1, UmiR167-2 to target ARF6/8 and non-ARFs respectively, is caused by their sequence variation in the course of evolution. Our recent study showed that miRNA such as miR166s, despite their sequence conservation, shows sequence variation in target complementary sites leading to the functional diversification in moss (Physcomitrella patens), where ppt-miR166m targets non-HD-ZIPIII transcript¹¹. As the sequence of UmiR167-2 was significantly different from UmiR167-1, it was not showing sufficient complementary binding with ARF6/8. The level of complementarity of a miRNA to its possible targets may also vary due to the sequence variation in the target sites. Therefore, we performed multiple sequence alignment between the targets MdCNBL10.2, 10.3 and 10.4 along with complementary sequence of UmiR167-2, which have shown 100% identity (Figure 1c). Further, AtCNBL10, an orthologue of MdCNBL10, was also included for multiple sequence alignment with UmiR167-2 and MdCNBL10.2, 10.3, 10.4. Inclusion of AtCNBL10 during multiple sequence alignment has again lowered

RESEARCH COMMUNICATIONS

the identity up to 55% (Figure 1 *d*). This suggests that the deviation in complementary miRNA binding site of a target gene depends upon the uniqueness of a miRNA as well as its target site sequences, which lead to the functional divergence. Subsequently, we observed that the UmiR167-2 targets, namely MdCNBL10.2, 10.3 and 10.4 encode for identical CNBL10 proteins. However, MdCNBL10.1, the closest homolog of AtCNBL10, codes for related protein but is not targeted by UmiR167-2. When the protein sequences of MdCNBL 10.2, 10.3, 10.4 and AtCNBL10 were aligned, it was found that amino acids at the miRNA binding site were not having 100% identity, having only 2 identical, 3 similar, and 1 less similar amino acid. This indicates that the target MdCNBL10 proteins have diverged from non-target MdCNBL10 proteins through sequence variation in critical miRNA complementary sites (Figure 1e). It implies that during the course of evolution, natural selection played a crucial role by choosing different targets among plant species for the same miRNA. The evolution of *MdCNBL10.2*, 10.3 and 10.4 as a target of mdm-miR167a is an interesting aspect to be understood. The cultivation of apple is done in cool temperate zones with warm days and cool nights, which resembles with cool, desert-like climate; the role of CNBL10 was reported as a calcium sensor, which was involved in the signalling pathway during salt and drought stresses¹². It is possible that the speciation and climatic adaptation in apple was contributed by the co-evolution of mdm-miR167a and its non-ARFs CNBL10 targets. Similarly, IAA-Ala Resistant3 (IAR3) was identified as a new target of ath-miR167a, which was cleaved during high osmotic stress^{13,14}. This indicates potential role of mdm-miR167a mediated regulation of MdCNBL10 during stress responses in apple.

Therefore, our findings shed light on the basis of functional diversification of miRNAs, as it is often believed that miRNAs and their targets are conserved. The deviation from the canonical target *ARFs* of a miR167 is due to altered pattern of processing of mdm-miR167a from the 3' end of the precursor sequences (instead of 5' end), which further led to the sequence variation in the mature miR167 targeting novel gene *MdCNBL10*.

- 3. Allen, E., Xie, Z., Gustafson, A. M. and Carrington, J. C., microRNA-directed phasing during trans-acting siRNA biogenesis in plants. *Cell*, 2005, **121**, 207–221.
- Llave, C., Xie, Z., Kasschau, K. D. and Carrington, J. C., Cleavage of Scarecrow-like mRNA targets directed by a class of *Arabidopsis* miRNA. *Science*, 2002, 297, 2053–2056.

- 5. Bonnet, E., Wuyts, J., Rouze, P. and Van de Peer, Y., Detection of 91 potential conserved plant microRNAs in *Arabidopsis thaliana* and *Oryza sativa* identifies important target genes. *Proc. Natl. Acad. Sci. USA*, 2004, **101**, 11511–11516.
- Barik, S. *et al.*, Coevolution pattern and functional conservation or divergence of miR167 and their targets across diverse plant species. *Sci. Rep.*, 2015, 5, doi:10.1038/srep14611.
- Wu, M. F., Tian, Q. and Reed, J. W., *Arabidopsis* microRNA167 controls patterns of *ARF6* and *ARF8* expression, and regulates both female and male reproduction. *Development*, 2006, 133, 4211–4218.
- Dai, X. and Zhao, P. X., psRNA target: a plant small RNA target analysis server. *Nucl. Acids Res.*, 2011, 39, W155–W159.
- Tamura, K., Stecher, G., Peterson, D., Filipski, A. and Kumar, S., MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol. Biol. Evol.*, 2013, **30**, 2725–2729.
- Larkin, M. A. et al., Clustal W and Clustal X version 2.0. Bioinformatics, 2007, 23, 2947–2948.
- Barik, S., Sarkar Das, S., Singh, A., Gautam, V., Kumar, P., Majee, M. and Sarkar, A. K., Phylogenetic analysis reveals conservation and diversification of micro RNA166 genes among diverse plant species. *Genomics*, 2014, **103**, 114–121.
- 12. Kim, B. G. *et al.*, The calcium sensor CBL10 mediates salt tolerance by regulating ion homeostasis in *Arabidopsis*. *Plant J.*, 2007, **52**, 473–484.
- Kinoshita, N. *et al.*, IAA-Ala Resistant3, an evolutionarily conserved target of miR167, mediates *Arabidopsis* root architecture changes during high osmotic stress. *Plant Cell*, 2012, 24, 3590– 3602.
- Sarkar, A. K., Karthikeyan, M., Gautam, V., Barik, S. and Das, S. S., Improving the plant root system architecture to combat abiotic stresses incurred by global climate changes. In *Climate Change* and Abiotic Stress Tolerance, Wiley, Germany, 2014, pp. 305– 324.

ACKNOWLEDGEMENTS. This work was supported by NIPGR short term fellowship, followed by SERB National Post-doctoral fellowship (PDF/2015/000232) to A.K. and NIPGR internal core grant to A.K.S.

Received 17 March 2016; revised accepted 10 August 2016

doi: 10.18520/cs/v112/i01/147-150

Singh, A., Singh, S., Panigrahi, K. C., Reski, R. and Sarkar, A. K., Balanced activity of microRNA166/165 and its target transcripts from the class III homeodomain leucine-zipper family regulates root growth in *Arabidopsis thaliana*. *Plant Cell Rep.*, 2014, 33(6), 945–953.

Arikit, S., Zhai, J. and Meyers, B. C., Biogenesis and function of rice small RNAs from non-coding RNA precursors. *Curr. Opin. Plant Biol.*, 2013, 16, 170–179.